



A *Talipariti elatus* Sw. Fryxell flowers extract inhibits histamine-induced edema in mice

Milagros García Mesa*, Abel Duménigo González, Lérica Lázara Acosta de la Luz, Yisel Blanco Hernández, Marisol López Barreiro

Central Laboratory of Pharmacology, Salvador Allende Faculty of Medicine, Medical University of Havana, Havana, Cuba

Correspondence to

Milagros García Mesa
Email:
milagros.mesa@infomed.sld.cu

Received 28 Dec. 2016

Accepted 20 Mar. 2017

ePublished 30 May 2017



Abstract

Introduction: *Talipariti elatus* Sw. Fryxell (Malvaceae) is a wooden botanical species that is endemic to Jamaica and Cuba. The main flavonoids identified in the flowers of this species have several biological activities, including the inhibition of vascular hyperpermeability induced by different stimuli. This work was aimed to assess whether a hydroalcoholic extract of *T. elatus* flowers (TFE) inhibits the edemagenic effect of histamine.

Methods: The flowers were collected from *T. elatus* trees at Cerro Municipality in Havana. TFE was obtained with 70% ethanol and was characterized by a phytochemical screening. Acute oral toxicity was determined in both sex SD rats. TFE (67; 200 and 600 mg/kg) or ketotifen (3 mg/kg) were orally administered to male Balb/c mice 1 hour before an intraplantar injection of histamine (50 µg) and the edemagenic reaction determined as the paw weight increment.

Results: Saponins, phenols- such as: tannins and flavonoids, anthocyanins, alkaloids, quinones, and coumarins were the major bioactive compounds in the extract. There were neither deaths nor any sign of toxicity among the rats treated with TFE. There was a significant inhibition of histamine-induced edema that was independent of TFE dose that was comparable with ketotifen.

Conclusion: This was the first evidence of the anti-edemagenic effect of the 70% ethanol extract of *T. elatus* flower petals against histamine-induced edema as a result of the early stage of a research project that deals with the pharmacological study of this endemic species.

Keywords: *Talipariti elatus*, Medicinal plant, Anti-edemagenic action, Histamine, Chemical composition, Acute toxicity

Introduction

Talipariti elatus Sw. Fryxell (majagua) fam. Malvaceae is a wooden botanical species that is endemic to Jamaica and Cuba (Figure 1).^{1,2} However, there is little information about the biological properties of this species.³

Nonclinical studies have provided evidence on some biological activities of quercetin, rutin, and gossypitrin, the major flavonoids of *T. elatus* flowers.⁴ For instance, quercetin and rutin inhibited histamine release from mast cells,^{5,6} reduced phospholipase A2 activity and recruitment of neutrophils and eosinophils into the lung.⁵ Furthermore, rutin is a known protector from capillary permeability by histamine and other compounds.^{7,8} Moreover, quercetin has induced the relaxation of airway smooth muscle, potentiated the β -agonist-induced effect on this tissue in vitro⁹ and inhibited the transcriptional up-regulation of histamine H1 receptor.¹⁰ Antioxidant capacity of the three flavonoids has been demonstrated.^{11,12} These biological properties could be transferred to a given product derived from this part of the plant. Therefore, a

research strategy for the characterization of its therapeutic potential should include the evaluation of its possible inhibitory effect on the release of chemical mediators, the contraction of airways smooth muscle and the increase of vascular permeability stimulated by histamine and other inflammatory agents. Accordingly, this study was aimed to assess the presence of one of these biological properties



Figure 1. *Talipariti elatus* Sw. Fryxell



in a hydro-alcohol extract of *T. elatus* flower petals; this is the ability to inhibit histamine-induced increase of capillary permeability. It is the first stage of a research project that deals with the pharmacological study of this endemic species.

Materials and Methods

Plant Material

Flowers were collected from *T. elatus* Sw. Fryxell trees planted in parterres at Cerro Municipality of Havana, in the morning during July and August 2015. The taxonomic identification of this botanic species was performed by the Central Laboratory of Pharmacology from Salvador Allende Faculty of Medicine, University of Medical Sciences of Havana, Cuba. Its identity was demonstrated by comparison with the sample registered at Havana Botanic Garden (Voucher number HAJB 82587). The flower petals were separated and cut into pieces, dried in an oven at 40°C for 12 hours (Figure 2), and reduced to small particles of about 3 mm by using a mesh battery.

Preparation of Plant Extract

An extract of *T. elatus* flower (TFE) petals was prepared by the method Miranda and Cuellar.¹³ It was obtained by percolation of dried plant material with 70% ethanol. Afterward, rotoevaporation (at 40°C and 27 mm Hg reduced pressure) of the homogeneous liquid obtained was performed for ethanol elimination from it in order to avoid the possible bias of the results of the pharmacological evaluations. The remaining liquid was stored in closed dark bottles at 4 to 8°C until used. The total soluble solids (TSS) content of the extract was gravimetrically determined in dried 1 mL aliquots (three replicates) by using a Sartorius MA40 analytical balance. A mean value of 400 mg/mL was obtained.

Phytochemical Screening

A phytochemical screening of TFE was performed by the method described earlier,¹³ that consisted of the formation of colored and/or insoluble chemical complexes from the secondary metabolites contained in the plant extract.

Animals

Sprague Dawley rats and Balb/c mice were purchased from the National Center for the Production of Laboratory Animals (Havana, Cuba). The animals were placed into polyurethane cages and housed at controlled environment (23°C and 50-60% relative humidity) with free access to food (AL- YCO® CMO 1000) and water.

Acute Oral Toxicity

TFE acute oral toxicity was evaluated in SD rats (both sexes, 200 ± 20 g b.w.) according to 423 OECD guideline.¹⁴ A single intragastric dose of the extract (2000 mg TSS/kg b.w.) was administered to rats. They were observed for a 14-day period in order to detect the occurrence of deaths



Figure 2. Drying of *T. elatus* flowers.

or any sign of toxicity. In addition, their body weights were determined before and after 7 and 14 days of treatment.

Increase of Histamine-Induced Vascular Hyper Permeability in the Mouse Paw

This assay was performed as described by Leal et al¹⁵ with modifications. Ten male Balb/c mice (20-25 g b.w.) were randomly assigned to each treatment group: negative control (saline), positive control (ketotifen 3 mg/kg b.w.) and TFE (67; 200 and 600 mg TSS /kg b.w.). Food was retired 12 hours before the experiment. Saline, ketotifen and TFE solutions (0.1 mL/30 g b.w.) respectively were intragastrically administered to the animals 1 hour before a subplantar injection of 50 µL of saline containing 50 µg of histamine into the right hind paw and the same volume of saline solution alone in the left hind paw. The animals were sacrificed by cervical dislocation 1 hour later and their paws were cut and weighted in Sartorius MA40 analytical balance. Results are expressed as the difference between right and left paw weight.

Statistical Analysis

The experimental results were expressed as the mean ± standard deviation (SD) of each group of treatment. Normal distribution of data and homogeneity of variance were assessed by Shapiro-Wilk's and Bartlett's tests, respectively. The statistical comparison between groups was performed by one-way ANOVA or Kruskal-Wallis test, according to the results of the previous tests. The means were compared by Dunn's and Bonferroni's tests. Differences lower than 5% were considered significant. The GraphPad Prism 5 statistical program was used for this purpose.

Ethics

All procedures described were carried out using a protocol approved by the Institutional Research Ethics Committee of Salvador Allende Faculty of Medicine. The guidelines for good practices for the management of laboratory animals^{16,17} were followed.

Results and Discussion

Phytochemical Screening of TFE

As can be observed in Table 1, saponins and phenols –

such as: tannins and flavonoids, anthocyanins, alkaloids, quinones, and coumarins – were the major secondary metabolites found in TFE, as previously reported for the flowers and ethanol extracts of *T. elatus* plants grown in Cuba.^{4,18,19} This qualitative similarity endorses the contribution of the results of this research for the chemical, pharmacological and toxicological characterization of this botanical species.

Oral Acute Toxicity

The demonstration that a research product has a low probability to be toxic for humans is pivotal for the strategy of new drug development.²⁰ This concept is valid for a plant extract, though it is obtained from parts of those called medicinal plants, since they have complex compositions, as demonstrated for TFE (Table 1), with the combination of different biological properties that not only could support their medicinal use, but also could result in undesirable bio- logical effects.

The evaluation of acute toxicity in experimental animals is a primary stage of non-clinical estimation of a product's safety. It consists of the evaluation of quantitative and qualitative changes, associated with the administration a single or repeated doses of the substance in a 24-hour period of time. This information offers a basis to calculate the doses of a product for its pharmacological characterization.

In agreement with a previous toxicological study of an ethanol extract of *T. elatus* flowers,²¹ the acute intragastric administration of TFE 2000 mgTSS/kg b.w to rats (a maximal allowable dose²⁰) caused neither deaths nor signs of toxicity among these animals during the 14-day period of observation following the treatment. Also, body weight gain was within the expected limits for this species (Figure 3), suggesting that this extract could be safe after a single dose treatment. However, other toxicological studies are necessary to complete the knowledge about this extract safety.

Table 1. Phytochemical Screening of TFE

Assay	Identified Metabolites	Results
Resins	Resinous products	–
Baljet	Lactonics compounds	++
Foam	Saponins	+++
Ferric chloride	Tannins and phenols	+++
Ninhydrine	Amino acids	–
Shinoda	Flavonoids	++
Kedde	Glycosides	–
Fheling	Reducing sugars	+++
Lieberman-Buchard	Triterpenes and steroids	+
Bortranger	Quinones	+++
Anthocyanidins	Anthocyanidins	+++
Drangendorff/Mayer /Wagner	Alkaloids	+++

+++ Strong reaction; ++ Intermediate reaction; + Weak reaction; - Absence of reaction.

Inhibition of the Increase of Histamine-Induced Capillary Permeability in the Mouse Paw

Preliminary experiments for standardization of this experimental model allowed the selection of the doses of histamine and the H1 antihistamine drug ketotifen, to be used in the experiments for the edemagenic stimulus and the positive control respectively. Fifty micrograms of histamine/paw showed to be a proper stimulus level because it produced a measurable, submaximal and reproducible effect. On the other hand, ketotifen 3 mg/kg b.w. had a submaximal inhibitory activity.

Since a single oral dose of TFE 2000 mg/kg showed no toxicity to animals a range of lower doses, being one 1/3 of the other (600; 200 and 67 mg/kg), was selected to assess the extract effect on histamine edemagenic activity in the mouse paw. These numbers approximately correspond to 1/3; 1/9 and 1/27 x 2000 mg/kg respectively.

Figure 4 shows that TFE significantly inhibited histamine induced edema at all dose levels tested but not in a dose-independent manner. Consequently, 67 mg/kg could be the highest effective dose of this extract and its ED 50 could be below it, thus suggesting that it has a favorable security index, higher than 27 after a single oral dose.

Histamine-induced paw edema was significantly lower in the group of mice treated with ketotifen with respect to the negative control group (Figure 4). The lack of statistical difference between the groups of animals treated with the extract and the one treated with the reference drug (Figure 4) is a demonstration of similar effectiveness in these experimental conditions that suggests a therapeutic potential of this plant preparation.

The flavonoids contained in TFE could be responsible for the inhibition of histamine effect but it has been

said that they do not have antihistamine activity.²² Consequently, it seems that an unspecific rather than a specific mechanism of action could explain the results. Nevertheless, a contribution of other chemical

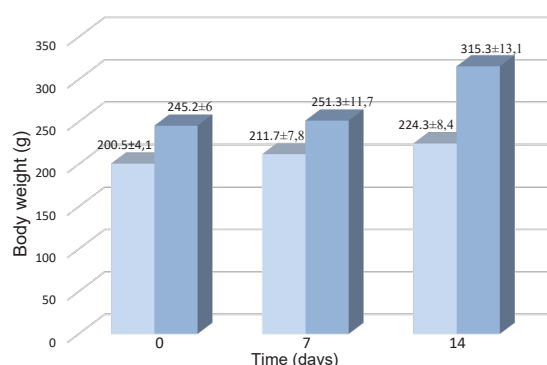


Figure 3. Increase of body weight in Wistar rats (female: gray and male: black) (N = 3 animals/group) after a single oral administration of TFE 2000 mg/kg b.w. One-way ANOVA with Bonferroni's post-test.

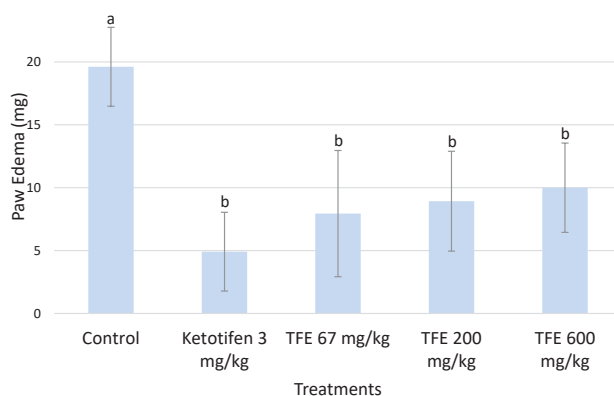


Figure 4. Antihistaminic effect of *Talipariti elatus* dried flowers fluid extract p.o. in Balb/c male mice (N = 10 animals/group). Kruskal–Wallis with Dunns’ post-test. Different letters means significant statistical differences for $P < 0.05$.

components to the biological activity of this extract should not be discarded. Additional studies should be aimed to clarify it, and its influence on vascular hyper permeability induced by other stimuli.

The results presented here support the hypothesis of this research that an extract of *T. elatus* flowers could inhibit the edemagenic effect of histamine. Nevertheless, its probable inhibitory effects on the release of histamine, leukotrienes and other inflammatory mediators, as well as on airway contraction and other biological activities associated with the main secondary metabolites present in *T. elatus* flowers should be evaluated.

Some chronic diseases involving edema and inflammation induced by histamine could also be influenced by this extract. For instance, rutin pharmaceutical products and the micronized purified flavonoid fraction have a demonstrated activity as venotonic agents and are used for the treatment of chronic venous insufficiency.²³⁻²⁵

Accordingly, this plant preparation could have a wide [i] field of therapeutic use that deserves further investigation.

Conclusion

The present study has given the first qualitative experimental evidence that a 70% ethanol extract of the dried flower petals of *T. elatus* plants grown in Cuba contains saponins and phenols such as tannins and flavonoids, anthocyanins, alkaloids, quinones, and coumarins. Also, it did not show acute toxicity in rats. Furthermore, it was able to inhibit histamine-induced paw edema in mice. It was a result of the early stage of a research project that deals with the pharmacological study of this endemic species.

Competing Interests

None.

References

1. Fuentes Fiallo VR, Exposito Montoya A. The ethnobotanical

surveys on medicinal plants in Cuba. *Rev Jard Bot Nac.* 1995;16:77-144.

2. Volpato G, Godinez D, Beyra A, Barreto A. Uses of medicinal plants by Haitian immigrants and their descendants in the province of Camaguey, Cuba. *J Ethnobiol Ethnomed.* 2009;5:16. doi:10.1186/1746-4269-5-16

3. Cuellar Cuellar A, Gonzalez Yaque JA. Isolation of the flavonoid glycoside gossypitrin from *Talipariti elatum* S.W. flowers and evaluation of its possible antioxidant effect. *RECIA.* 2010;2(2):338-348.

4. Marquez Hernandez I, Cuellar Cuellar A, Martinez Perez J, Aleman Sanchez A, Lora Garcia J, Velez Castro H. Phytochemical study of the species *Hibiscus elatus* SW. *Rev Cubana Farm.* 1999;33(2):127-131.

5. Jung CH, Lee JY, Cho CH, Kim CJ. Anti-asthmatic action of quercetin and rutin in conscious guinea-pigs challenged with aerosolized ovalbumin. *Arch Pharm Res.* 2007;30(12):1599-1607. doi:10.1007/bf02977330

6. Kempuraj D, Madhappan B, Christodoulou S, et al. Flavonols inhibit proinflammatory mediator release, intracellular calcium ion levels and protein kinase C theta phosphorylation in human mast cells. *Br J Pharmacol.* 2005;145(7):934-944. doi:10.1038/sj.bjp.0706246

7. Benavente-Garcia O, Castillo J, Marin FR, Ortuno A, Del Rio JA. Uses and Properties of Citrus Flavonoids. *J Agric Food Chem.* 1997;45(12):4505-4515. doi:10.1021/jf970373s

8. Benavente-Garcia O, Castillo J. Update on uses and properties of citrus flavonoids: new findings in anticancer, cardiovascular, and anti-inflammatory activity. *J Agric Food Chem.* 2008;56(15):6185-6205. doi:10.1021/jf8006568

9. Townsend EA, Emala CW Sr. Quercetin acutely relaxes airway smooth muscle and potentiates beta-agonist-induced relaxation via dual phosphodiesterase inhibition of PLCbeta and PDE4. *Am J Physiol Lung Cell Mol Physiol.* 2013;305(5):L396-403. doi:10.1152/ajplung.00125.2013

10. Hattori M, Mizuguchi H, Baba Y, et al. Quercetin inhibits transcriptional up-regulation of histamine H1 receptor via suppressing protein kinase C-delta/extracellular signal-regulated kinase/poly(ADP-ribose) polymerase-1 signaling pathway in HeLa cells. *Int Immunopharmacol.* 2013;15(2):232-239. doi:10.1016/j.intimp.2012.12.030

11. Lago JH, Toledo-Arruda AC, Mernak M, et al. Structure-activity association of flavonoids in lung diseases. *Molecules.* 2014;19(3):3570-3595. doi:10.3390/molecules19033570

12. Perez-Trueba G, Ramos-Guanche C, Martinez-Sanchez B, Marquez-Hernandez I, Giuliani A, Martinez-Sanchez G. Protective effect of gossypitrin on carbon tetrachloride-induced in vivo hepatotoxicity. *Redox Rep.* 2003;8(4):215-221. doi:10.1179/135100003225002718

13. Miranda M, Cuellar A. Pharmacognosy and natural products. Havana, Cuba: Editorial Felix Varela; 2001.

14. Acute Toxic Class Method. OECD (Organization for Economic Cooperation and Development) Guideline 423;2001.

15. Leal LK, Canuto KM, da Silva Costa KC, et al. Effects of amburoside A and isokaempferide, polyphenols from *Amburana cearensis*, on rodent inflammatory processes and myeloperoxidase activity in human neutrophils. *Basic Clin Pharmacol Toxicol.* 2009;104(3):198-205. doi:10.1111/j.1742-7843.2008.00329.x

16. National Research Council. Guide for the Care and Use of Laboratory Animals. Washington DC, EEUU: National Academies Press; 2001:21-79.

17. For Public Health Protection. The bases for Sanitary and Environmental Good Practices for Non-Clinical Laboratories. Havana, Cuba: Bureau of Regulatory Affairs; 2004.

18. Milanés Santana R, Alonso Rodríguez D, González Aguilar

- G. Pharmacognostic study of "majagua flowers" plant drug (*Hibiscus elatus* Sw., Malvaceae family). III. Crude drug standardization. Drug alterations. *Rev Cub Plant Med.* 1999;4(2):79-81.
19. Milanes Santana R, Alonso Rodriguez D, Gonzalez Aguilar G. Pharmacognostic study of "majagua flowers" plant drug (*Hibiscus elatus* Sw., Malvaceae family). IV. Fluid extract standardization. *Rev Cub Plant Med.* 1999;4(2):82-87.
20. Dispasquale L, Wallace H. Acute toxicity and eye irritancy. In: Wallace H, ed. Principles and methods of toxicology. 4th ed. Philadelphia: Taylor and Francis; 2001:853-917.
21. Monteagudo Jimenez EE, Boffil Cardenas MA, Verdecia Machado B. Acute toxicity of six Cuban medicinal plants by acute class toxicity method. *Medicentro.* 2007;11(1).
22. Wilson RH, Booth AN, DeEds F. Protection by flavonoids against histamine shock. *Exp Biol Med.* 1951;76(3):540-542.
23. Clement DL. Management of Venous Edema: Insights from an International Task Force. *Angiology.* 2000;51(1):13-17. doi:10.1177/000331970005100104
24. Frick RW. Three treatments for chronic venous insufficiency: escin, hydroxyethylrutoside, and Daflon. *Angiology.* 2000;51(3):197-205. doi:10.1177/000331970005100303
25. Katsenis K. Micronized purified flavonoid fraction (MPFF): a review of its pharmacological effects, therapeutic efficacy and benefits in the management of chronic venous insufficiency. *Curr Vasc Pharmacol.* 2005;3(1):1-9. doi:10.2174/1570161052773870

© 2017 The Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.